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学 位 論 文

Differential regulation and correlation between galectin-9 and
anti-CCP antibody in rheumatoid arthritis patients

(関節リウマチ患者におけるガレクチン9と抗CCP抗体の相関と
病態形成への関与)

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Background

Galectin-9 (Gal-9) is involved in the regulatory process of immune responses or inflammation. The inhibitory effect of Gal-9 is expected to the relation of pathophysiology in many autoimmune diseases. In collagen-induced arthritis model mice, administration of Gal-9 has been reported to reduce arthritis. Therefore, Gal-9 could be an important regulator of RA. The aim of the present study is to characterize circulating Gal-9 in patients with rheumatoid arthritis (RA) and its relationship with RA disease activity and phenotype.

Methods

A total of 116 RA patients and 31 age-matched healthy controls were included in this study. Disease activity of RA patients was determined by Disease Activity Score of 28 joint scoring system (DAS28-ESR). Levels of Gal-9 in serum were determined by enzyme-linked immunosorbent assay (ELISA).

Results

Serum levels of Gal-9 were significantly higher in patients with RA compared to those in controls (median 7577 pg/ml [interquartile range (IQR) 5570–10,201] versus 4738 pg/ml [IQR 4267–5630], $p = 0.001$). There were significant differences in serum Gal-9 between with and without advanced joint damage (8790 pg/mL [IQR 5631–10953] versus 7103 pg/mL [IQR 5882–8810], $p < 0.023$). Although serum levels of Gal-9 correlated with the titers of anti-CCP antibody (ACPA) ($r = 0.275$, $p = 0.002$), levels of ACPA titers conferred the different relationship, between serum Gal-9 and inflammatory mediators or RA disease activity. Although Gal-9 was correlated with ACPA titers ($r = 0.508$, $p = 0.002$), there was no correlation between Gal-9 levels and erythrocyte sedimentation rate (ESR), matrix metalloproteinase-3 (MMP-3), or DAS28-ESR in RA patients with high titers of ACPA (> 200 U/ml). Conversely, Gal-9 was correlated with MMP-3 ($r = 0.300$, $p = 0.007$) or DAS28-ESR ($r = 0.331$, $p = 0.004$) but not with ACPA titer in RA patients with low titers of ACPA (< 200 U/ml).

Conclusions

The levels of ACPA titers may influence the values of circulating Gal-9 in RA patients with various clinical phenotypes. Therefore, the combination of ACPA titers and Gal-9 may be useful for classifying the pathophysiology of RA. These data suggest that Gal-9 possessed the properties of pro-inflammatory or arthropathic biomarker under the status of ACPA titers

Introduction

Rheumatoid arthritis (RA) is an autoimmune disease characterized by synovial inflammation, bone destruction, and extra-articular symptoms [1]. Multiple genes, proteins, and cells have been identified to contribute to the pathogenesis of RA [2]. A common feature of RA is the hyper-activated state of the stromal synovial cells and the immune cells [3]. Dysregulated innate and adaptive immunities are involved in the RA pathogenesis [4]. Galectins are the family members of lectins that expressed on the cell surface or extracellular matrix and bind to β -galactoside carbohydrates on the cell surface [5]. Through binding to their receptors, galectins play an important role in the pathological processes including inflammation and autoimmunity [6]. Recent studies suggest that galectins play important roles in the pathogenesis of RA [7]. It was demonstrated that galectin-3 is increased in early RA and associated with anti-CCP seropositivity and MRI bone erosion scores in patients with RA [8]. These findings suggest that galectin family plays an important role in the disease development of RA, through their interactions with innate or adaptive immunity.

Gal-9 is expressed by immune cells, endothelial cells, and fibroblasts and plays an important role in regulating inflammation and immune reactions [9]. Gal-9 is a ligand of T cell immunoglobulin and mucin-containing-molecule-3 (Tim-3) which is expressed on CD4⁺ T helper (Th) 1 and Th17 and providing inhibitory signals through its interaction with Tim-3 [10]. Therefore, Gal-9 negatively regulates pro-inflammatory T cell responses through the interaction with Tim-3 and Gal-9/Tim-3 pathway induces apoptosis of CD4⁺ Th1 or Th17 cells [11]. In addition, the function of Tim-3 affects myeloid cells such as monocytes, macrophages, and dendritic cells, resulting in regulation of cytokine production [12]. In mouse models, Gal-9 deficiency led to increased number of Th1 and Th17 cells and decreased number of Treg cells in the joint, rendering susceptibility to collagen-induced arthritis (CIA) [13]. Conversely, administration of Gal-9 ameliorated arthritis in CIA and immune complex-induced murine arthritis model suggesting that Gal-9 prevents the disease progression of RA [14]. Considering that RA is regarded as a Th1-polarized autoimmune disease, dysregulated Gal-9 levels may cause the imbalance in the innate/adaptive immunity, thereby inducing pathological rheumatoid inflammation. In inflammatory arthritis, Gal-9 was shown to mediate the angiogenesis and infiltrations of inflammatory cells [15]. These findings suggest that Gal-9 may contribute to the rheumatoid inflammatory processes. Therefore, we focused on Gal-9 and hypothesized that Gal-9 may play a role in the pathogenesis of RA. In this study, we examined the levels of serum Gal-9 in

patients with RA and evaluated the results with respect to the clinical parameters.

Methods

Patients

A total of 136 patients diagnosed with RA between February 2012 and September 2019 were recruited from the Department of Rheumatology, Fukushima Medical University. Twenty RA patients complicated with infection, malignancy and other rheumatic diseases were excluded. All patients were fulfilled the 2010 ACR/EULAR classification criteria for the disease [16]. Finally, 116 consecutive RA patients were included in this study. We retrospectively reviewed the records of these RA patients. All patients were treated in the Department of Rheumatology, Fukushima Medical University, from June 2009 to March 2019.

The following clinico-demographic data were collected from the Medical Records Unit at Fukushima University Hospital: age, age at onset of RA, gender, Disease Activity Score-28 for Rheumatoid Arthritis with erythrocyte sedimentation rate (ESR) (DAS28-ESR) score [17], and extra-articular manifestations. This study was conducted in accordance with the principles of the Declaration of Helsinki. Ethical approval for this study (no. 2019097) was provided by the Ethics Committee of Fukushima Medical University.

Measurement of clinical disease activity

All patients underwent clinical assessment at baseline, including 28-joint swollen and tender joint counts (28-SJC and 28-TJC, respectively), physician and patient global assessment with visual analogue scales (0–100 mm), and ESR (mm). The composite disease activity indices were subsequently calculated: DAS28-ESR [16]. Result of this score was reported in quantitative value divided into 4 categories: remission with score of < 2.6 , mild activity if score of ≥ 2.6 to < 3.2 , moderate activity if score of ≥ 3.2 to < 5.1 , and high activity if score of ≥ 5.1 . Serum MMP-3 levels were measured by latex immunoassay (Panaclear MMP-3 “Latex”; Sekisui Medical Company Limited, Tokyo, Japan). The patients’ anti-CCP antibodies (ACPA) were analyzed using commercially available second-generation chemiluminescent enzyme immunoassay kits (STACIA® MEBLux™ CCP test, Medical and Biological Laboratories, Aichi, Japan) according to the manufacturer’s instructions. The results were reported qualitatively where negative or positive for ACPA was defined as < 20.0 U/ml or ≥ 20.0 U/ml, respectively. Radiographs were taken of both hands of each patient. Two rheumatologists, blinded to

the patient's identity and functional status, independently graded each hand radiographs and assigned as Steinbrocker radiographic stage [18].

ELISA methods

Serum concentrations of galectin-9 were measured using enzyme-linked immunosorbent assay kit (R&D Systems, Minneapolis, MN, USA) according to the manufacturer's instruction.

Statistical analysis

Results were non-normally distributed and are presented throughout the manuscript with median and 25–75th centiles (median, interquartile range [IQR]) and were compared by the Mann-Whitney U test. Correlations between continuous variables were analyzed by Spearman's rank correlation test. All data entry and statistical analyses were performed using SPSS Statistics version 22.0 (IBM, Armonk, NY). In all the analyses, a 2-tailed $p < 0.05$ was considered statistically significant.

Results

We recruited 116 patients with RA and 31 gender- and age-matched healthy subjects. The demographics and clinical characteristics of the RA patients are presented in Table 1. Among 116 patients, 83 patients (71.6%) were female. The median age of RA patients was 66 years. The majority of RA patients received DMARDs, mostly methotrexate or MTX in combination with other synthetic DMARDs. Despite the treatments, there was a median DAS28-ESR score of 2.8.

We measured serum levels of Gal-9 in RA patients using a specific ELISA assay. As shown in Fig. 1, serum Gal-9 concentrations in patients with RA were significantly higher compared to those in healthy subjects (median 7577 pg/ml [interquartile range (IQR) 5570–10,201] versus 4738 pg/ml [IQR 4267–5630], $p = 0.001$). We investigated the relationship between serum Gal-9 and each parameter of RA patients. Serum Gal-9 were significantly correlated with ESR ($r = 0.344$, $p < 0.001$), MMP-3 ($r = 0.234$, $p = 0.004$), and DAS28-ESR ($r = 0.269$, $p = 0.005$). Although we investigated the correlation between rheumatoid factor and serum Gal-9, there was no significant correlation between rheumatoid factor and Gal-9 ($r = 0.16$, $p = 0.09$). The positive correlation between Gal-9 and ACPA titer was observed ($r = 0.275$, $p = 0.002$, Fig. 2A). In the scatter plot, the distribution of Gal-9 was different depending on the level of ACPA. We subdivided into two groups, ACPA high titer group (ACPA ≥ 200 U/mL) and low titer group (ACPA < 200 U/mL). The correlation between Gal-9 and ACPA titer

in patients with high titers of ACPA was stronger than in patients with low titers of ACPA (Figure 2B). The clinical parameters also tended to differ between the two groups. In patients with low titers of ACPA, Gal-9 had positive correlation with ESR ($r = 0.451$, $p < 0.001$, Fig. 3A), DAS28-ESR ($r=0.331$, $p=0.004$, Fig. 3B) and MMP ($r = 0.300$, $p = 0.007$, Fig. 3C). In contrast, there were no correlations among Gal-9 and these parameters in patients with high titers of ACPA.

From a clinical point of view, the circulating Gal-9 were compared according to the presence or absence of clinical remission (CR) in these subgrouped RA patients (Fig. 4). In RA patients with low titers of ACPA (< 200 U/ml), circulating Gal-9 were significantly higher in patients without CR compared to those with clinical remission (8252 pg/ml [IQR 5870–10,996] versus 7103 pg/ml [IQR 5328–8357], $p = 0.013$). There was no significant difference in circulating Gal-9 between patients with and without CR (10,647 pg/ml [IQR 6960–13,367] versus 8635 pg/ml [IQR 6372–10,092], $p = 0.703$) in RA patients with high titers of ACPA (≥ 200 U/ml). Finally, we subdivided RA patients according to the progressing of joint damage (stage) and we evaluated the relationship between serum Gal-9 and progressive joint damage. As shown in Fig. 5A, RA patients with advanced articular lesions (stage II–IV) had significantly higher levels of circulating Gal-9 compared to those without advanced articular lesions (stage I) (8790 pg/ml [IQR 5631–10,953] versus 7103 pg/ml [IQR 5882–8810], $p = 0.023$). There was a significant difference in circulating Gal-9 between those with and without advanced articular lesions (stage II–IV) in RA patients with low titers of ACPA (7367 pg/ml [IQR 5931–10,109] versus 7009 pg/ml [IQR 4602–9450], $p = 0.004$). In contrast, there was no significant difference in circulating Gal-9 between those with and without advanced articular lesions (stage II–IV) in RA patients with high titers of ACPA (10,146 pg/ml [IQR 8771–12,257] versus 5882 pg/ml [IQR 5053–8138], $p = 0.182$) (Fig. 5B).

Discussion

In this study, we evaluated circulating levels of Gal-9 in established RA patients with various disease activities. We demonstrated that Gal-9 is significantly elevated in RA patients and correlated with the titers of ACPA as well as rheumatoid inflammatory markers such as ESR or MMP-3. Conversely, we could not find positive correlations between Gal-9 concentrations and MMP-3 or DAS-28-ESR in sub-grouped RA patients with high titers of ACPA (> 200 U/ml). These results suggest that galectin-9 could be a biomarker estimating RA disease activity under the particular ACPA status.

Variation of disease course and treatment response in RA patients originate from the heterogeneity of this syndrome [27]. There is evidence that the pathogenesis of RA differs between ACPA-positive and ACPA-negative RA patients [19]. This antibody could be used as a prognostic factor, since high titers for ACPA are associated with worse radiographic progression [20]. It was also postulated that an association between seropositivity of ACPA and response to abatacept (ABT) could be caused by blocking the interaction between T cells and APC via costimulatory molecules in which the effects of ABT on T cell co-stimulation are more marked in ACPA-positive patients [21]. In addition, the presence or absence of ACPA is also affect the cytokine profiles of RA [28]. These data suggest that RA patients with high titer of ACPA represent a unique RA population.

Gal-9 has been suggested to play a role in RA pathogenesis, but the underlying mechanisms have not been elucidated. Previous studies suggest that the measurement of disease activity alone is not sufficient to identify fast progressing RA, and high titers of ACPA could be a risk factor for RA progression [22, 23]. The correlation between Gal-9 and ACPA observed in our study may suggest that high titers of ACPA may be linked to the rheumatoid inflammatory process through the Gal-9-mediated bone effects indirectly. Interestingly, we found increased serum levels of Gal-9 in RA patients with progressive joint damage. Galectin family is involved in the rheumatoid osteoclastogenesis and inflammatory bone destruction [24]. Expression of Tim-3 was demonstrated in osteoclasts, and Gal-9 markedly inhibited osteoclastogenesis via Tim-3/Gal-9 system [25]. Therefore, it is possible that the Tim-3/Gal-9 system may regulate the rheumatoid inflammatory bone destruction. Taken together, the elevated circulating levels of Gal-9 seen in RA patients may reflect the augmented status of osteoclastogenesis of RA joints. It was reported that unlike to exogenous Gal-9, endogenous Gal-9 is protective against apoptosis and enhances synovial fibroblast viability suggesting the pathogenic and pro-inflammatory role in RA [26]. Considering the complex biological role of Gal-9, further studies are needed to determine the mechanism for the increased Gal-9 expression and its function upon the disease activity and joint damage of RA patients.

Sub-grouped RA patients with low titers of ACPA (< 200 U/ml) is especially related to inflammatory marker and disease activity score, such as DAS-28 or MMP-3. Tim-3 is also expressed on innate immune cells as well as adaptive immune cells. Tim-3 is one of the inflammatory regulators in macrophages. Macrophage are known to be involved in

the progression and inflammation of RA. Therefore, the dysfunction of Gal-9/TIM-3 system may result in the active inflammation in specifically subset of RA.

From these findings, it is also presumed that stratifying patients with RA on the basis of ACPA and Gal-9 status enables to identify more homogenous RA phenotype with respect to disease activity or joint structure damages.

There are several potential limitations of this study that should be considered. First, the patient population was relatively small and a larger study is essential to confirm our results. Second, all patients with RA and healthy individuals in this study were Japanese; additional studies in other ethnic groups are needed to verify these findings. Third, the mechanism by which Gal-9 contributes to the pathogenesis of RA was not clarified. Finally, it will be important to examine the longitudinal changes of serum galectin-9 levels in patients with RA and to assess their extra-articular involvements in the future studies. Nevertheless, our findings suggest that Gal-9 may be involved in the pathophysiology of RA reflecting disease activity or immune phenotypes of RA.

Conclusions

Serum Gal-9 shows as an additional biomarker for evaluating disease activity in patients with RA. Prospective investigation of the combination of Gal-9 and ACPA may facilitate development of diagnostic tools to assess disease activity and disease phenotype in patients with RA.

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References

1. Smolen JS, Aletaha D, McInnes IB. Rheumatoid arthritis. *Lancet*. 2016;388(10055):2023-2038.
2. Jutley G Raza K, Buckley CD. New pathogenic insights into rheumatoid arthritis. *Curr Opin Rheumatol*. 2015;27(3):249-55.
3. Bartok B, Firestein GS. Fibroblast-like synoviocytes: key effector cells in rheumatoid arthritis. *Immunol Rev*. 2010;233(1):233-55.
4. Catrina AI, Joshua V, Klareskog L, Malmström V. Mechanisms involved in triggering rheumatoid arthritis. *Immunol Rev*. 2016;269(1):162-74.
5. Thiemann S, Baum LG. Galectins and Immune Responses-Just How Do They Do Those Things They Do? *Annu Rev Immunol*. 2016;34:243-64.
6. Rabinovich GA, Rubinstein N, Toscano MA. Role of galectins in inflammatory and immunomodulatory processes. *Biochim Biophys Acta*. 2002;1572(2-3):274-84.
7. Ilarregui JM, Bianco GA, Toscano MA, Rabinovich GA. The coming of age of galectins as immunomodulatory agents: impact of these carbohydrate binding proteins in T cell physiology and chronic inflammatory disorders. *Ann Rheum Dis*. 2005;64 Suppl 4:iv96-103.
8. Issa SF, Christensen AF, Lindegaard HM, et al. Galectin-3 is Persistently Increased in Early Rheumatoid Arthritis (RA) and Associates with Anti-CCP Seropositivity and MRI Bone Lesions, While Early Fibrosis Markers Correlate with Disease Activity. *Scand J Immunol*. 2017;86(6):471-478.
9. Gieseke F, Kruchen A, Tzaribachev N, Bentzien F, Dominici M, Müller I. Proinflammatory stimuli induce galectin-9 in human mesenchymal stromal cells to suppress T-cell proliferation. *Eur J Immunol*. 2013;43(10):2741-9.
10. Zhu C, Anderson AC, Kuchroo VK. TIM-3 and its regulatory role in immune responses. *Curr Top Microbiol Immunol*. 2011;350:1-15.
11. Zhu C, Anderson AC, Schubart A, et al. The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. *Nat Immunol*. 2005;6(12):1245-52.
12. Ocaña-Guzman R, Torre-Bouscoulet L and Sada-Ovalle I. TIM-3 regulates distinct functions in macrophages. *Front. Immunol*. 2016;7: 229.
13. Arikawa T, Watanabe K, Seki M, et al. Galectin-9 ameliorates immune complex-induced arthritis by regulating Fc gamma R expression on macrophages. *Clin Immunol*. 2009;133(3):382-92.
14. Seki M, Sakata KM, Oomizu S, et al. Beneficial effect of galectin 9 on rheumatoid arthritis by induction of apoptosis of synovial fibroblasts. *Arthritis Rheum*. 2007;56(12):3968-76.

15. O'Brien MJ, Shu Q, Stinson WA, Tsou PS, Ruth JH, Isozaki T, Campbell PL, Ohara RA, Koch AE, Fox DA, Amin MA. A unique role for galectin-9 in angiogenesis and inflammatory arthritis. *Arthritis Res Ther.* 2018;20(1):31.
16. van der Linden MP, Knevel R, Huizinga TW, van der Helm-van Mil AH. Classification of rheumatoid arthritis: comparison of the 1987 American College of Rheumatology criteria and the 2010 American College of Rheumatology/European League Against Rheumatism criteria. *Arthritis Rheum.* 2011;63(1):37-42.
17. Fransen J, van Riel PL. The Disease Activity Score and the EULAR response criteria. *Rheum Dis Clin North Am.* 2009 Nov;35(4):745-57, vii-viii.
18. Steinbrocker O, Traeger CH, Batterman RC. Therapeutic criteria in rheumatoid arthritis. *J Am Med Assoc.* 1949;140:659-62.
19. Willemze A, Trouw LA, Toes RE, Huizinga TW. The influence of ACPA status and characteristics on the course of RA. *Nat Rev Rheumatol.* 2012;8(3):144-52.
20. Degboé Y, Constantin A, Nigon D, et al. Predictive value of autoantibodies from anti-CCP2, anti-MCV and anti-human citrullinated fibrinogen tests, in early rheumatoid arthritis patients with rapid radiographic progression at 1 year: results from the ESPOIR cohort. *RMD Open.* 2015;1(1):e000180.
21. Sokolove J, Schiff M, Fleischmann R, et al. Impact of baseline anti-cyclic citrullinated peptide-2 antibody concentration on efficacy outcomes following treatment with subcutaneous abatacept or adalimumab: 2-year results from the AMPLE trial. *Ann Rheum Dis.* 2016;75(4):709-14.
22. Sewerin P, Vordenbaeumen S, Hoyer A, et al. Silent progression in patients with rheumatoid arthritis: is DAS28 remission an insufficient goal in RA? Results from the German Remission-plus cohort. *BMC Musculoskelet Disord.* 2017;18:163.
23. Steffen U, Schett G, Bozec A. How Autoantibodies Regulate Osteoclast Induced Bone Loss in Rheumatoid Arthritis. *Front Immunol.* 2019;10:1483.
24. Moriyama K, Kukita A, Li YJ, et al. Regulation of osteoclastogenesis through Tim-3: possible involvement of the Tim-3/galectin-9 system in the modulation of inflammatory bone destruction. *Lab Invest.* 2014;94(11):1200-11.
25. Li YJ, Kukita A, Teramachi J, et al. A possible suppressive role of galectin-3 in upregulated osteoclastogenesis accompanying adjuvant-induced arthritis in rats. *Lab Invest.* 2009;89(1):26-37.
26. Pearson MJ, Bik MA, Ospelt C, et al. Endogenous Galectin-9 Suppresses Apoptosis in Human Rheumatoid Arthritis Synovial Fibroblasts. *Sci Rep.* 2018;8(1):12887.
27. England BR, Thiele GM, Mikuls TR. Anticitrullinated protein antibodies: origin and role in the pathogenesis of rheumatoid arthritis. *Curr Opin Rheumatol.*

2017;29(1):57-64.

28. Ridgley LA, Anderson AE, Pratt AG. What are the dominant cytokines in early rheumatoid arthritis? *Curr Opin Rheumatol.* 2018;30(2):207-214.

Figures and Tables

Table 1. Baseline characteristics of 116 Japanese patients with RA

Characteristics	Value
Age (years), median (IQR)	66 (56-73)
Female, n (%)	83 (71.6)
Smoker, n (%)	44 (37.9)
RA-ILD, n (%)	31 (26.7)
Duration of RA (year), median (IQR)	5 (2-10)
ESR (mm/h), median (IQR)	15.5 (7-27)
CRP (mg/dL), median (IQR)	0.29 (0.09-0.9)
MMP-3 (ng/mL), median (IQR)	106 (61.8-207.3)
RF (IU/mL), median (IQR)	44 (11.8-149.5)
Anti CCP-Ab (U/mL), median (IQR)	60.1 (4.0-373.1)
Corticosteroid, n (%)	53 (45.7)
Methotrexate, n (%)	59 (50.9)
Biologics, n (%)	38 (32.8)
DAS28-ESR, median (IQR)	2.8 (2.0-3.7)
Steinbrocker stage	I : 35, II :40, III 26, IV 13

ILD=interstitial lung disease, ESR=erythrocyte sedimentation rate, CRP=C reactive protein, MMP-3=matrix metalloproteinase-3, RF=rheumatoid factor, CDAI=Clinical Disease Activity index, SDAI=simplified disease activity index, DAS28=Disease Activity Score, IQR=interquartile range

Figure 1. Serum levels of galectin-9 in RA patients and controls.

Serum levels of galectin-9 in RA patients (n = 116) were significantly higher compared to those in healthy subjects (n = 31)

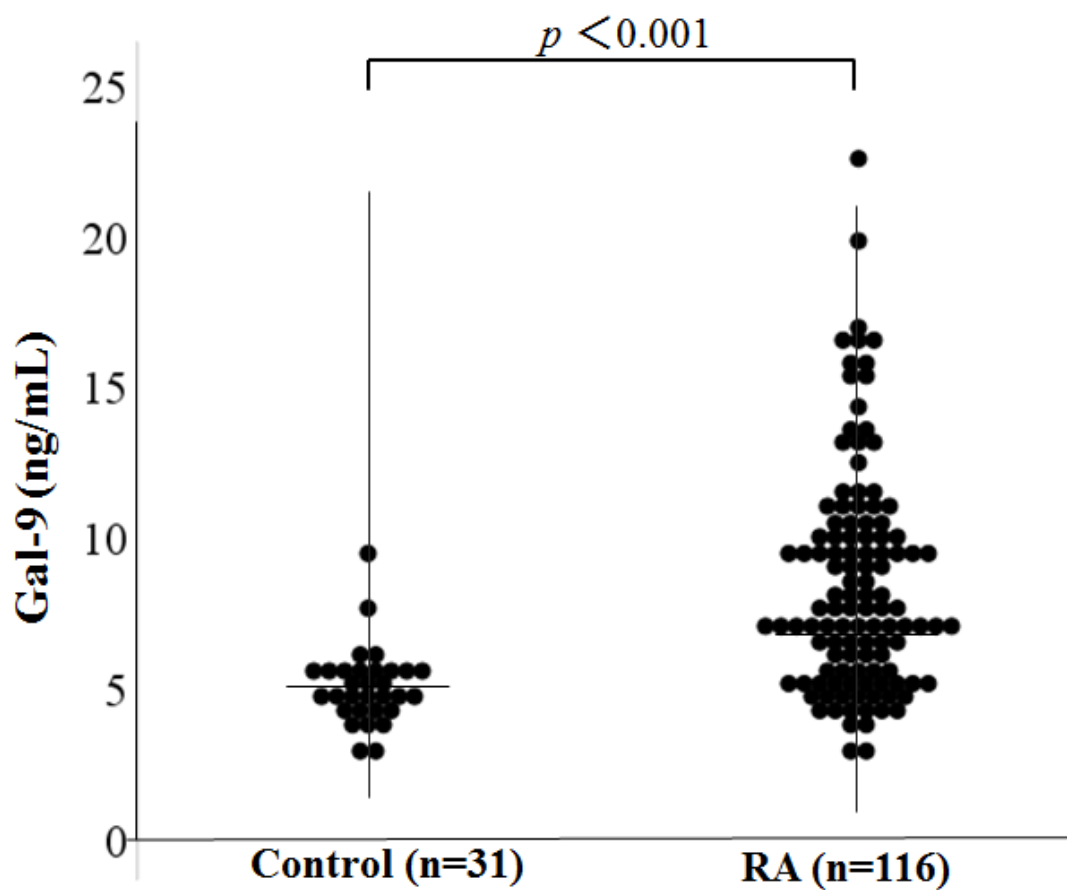


Figure 2. Relationship between ACPA titers and serum levels of galectin-9 in patients with RA.

A. Levels of ACPA titers were measured, and correlation analysis with serum levels of galectin-9 was performed.

B. Correlation analysis of serum levels of galectin-9 and ACPA titers does not show a relationship in RA patients with low titers of ACPA (< 200 U/ml), whereas there was a significant positive correlation between serum levels of Gal-9 and ACPA titers in RA patients with high titers of ACPA (≥ 200 U/ml)

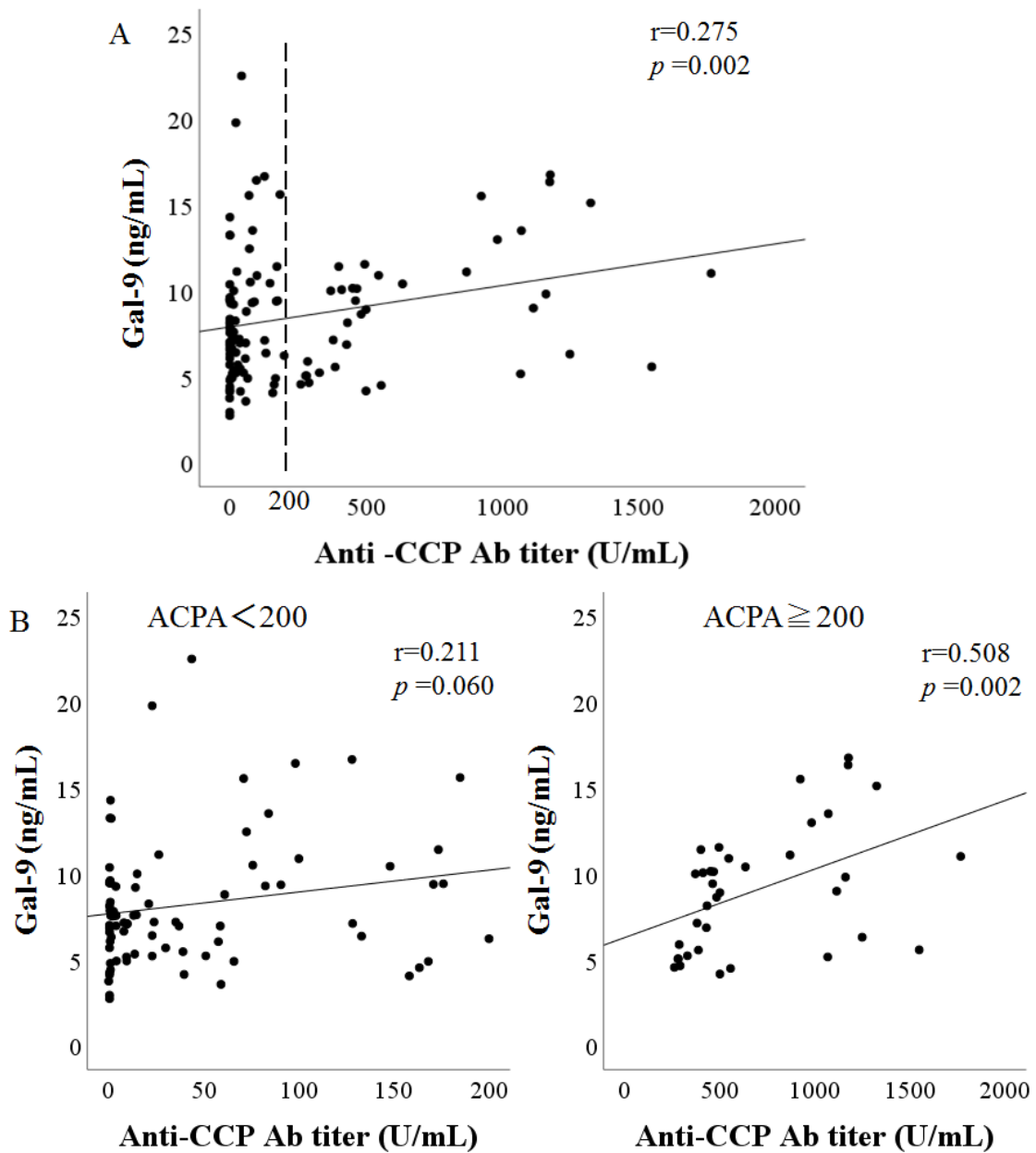


Figure 3. Correlation between serum levels of galectin-9 and clinical parameters in the sub-grouped RA patients according to the titers of ACPA.

A. Correlation analysis of serum levels of galectin-9 and ESR does not show a relationship in group 1 RA patients with high titers of ACPA (≥ 200 U/ml), whereas there was a significant positive correlation between serum levels of galectin-9 and ESR in group 2 RA patients with low titers of ACPA (< 200 U/ml).

B. Correlation analysis of serum levels of galectin-9 and DAS28-ESR does not show a relationship in group 1 RA patients with high titers of ACPA (≥ 200 U/ml), whereas there was a significant positive correlation between serum levels of galectin-9 and DAS28-ESR in group 2 RA patients with low titers of ACPA (< 200 U/ml).

C. Correlation analysis of serum levels of galectin-9 and MMP-3 does not show a relationship in group 1 RA patients with high titers of ACPA (≥ 200 U/ml), whereas there was a significant positive correlation between serum levels of galectin-9 and MMP-3 in group 2 RA patients with low titers of ACPA (< 200 U/ml).

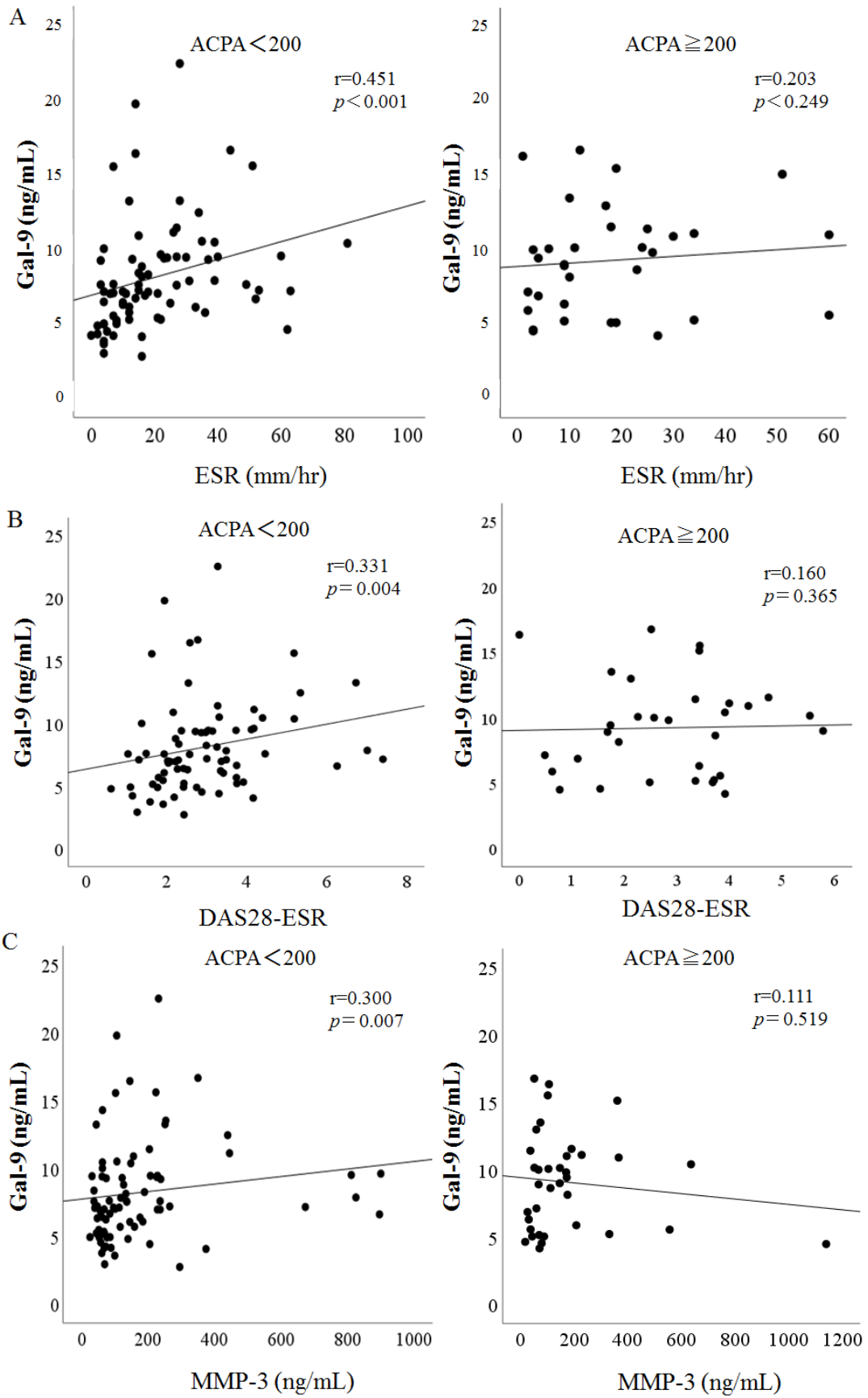


Figure 4. Serum levels of galectin-9 in RA patients with or without DAS28-ESR clinical remission (CR).

We compared serum levels of galectin-9 between RA patients with or without clinical remission (CR). Serum levels of galectin-9 were significantly lower in patients with CR compared to those without CR in RA patients with low titers of ACPA (< 200 U/ml), whereas there was no significant difference in serum levels of galectin-9 between patients with and without CR in RA patients with high titers of ACPA (≥ 200 U/ml).

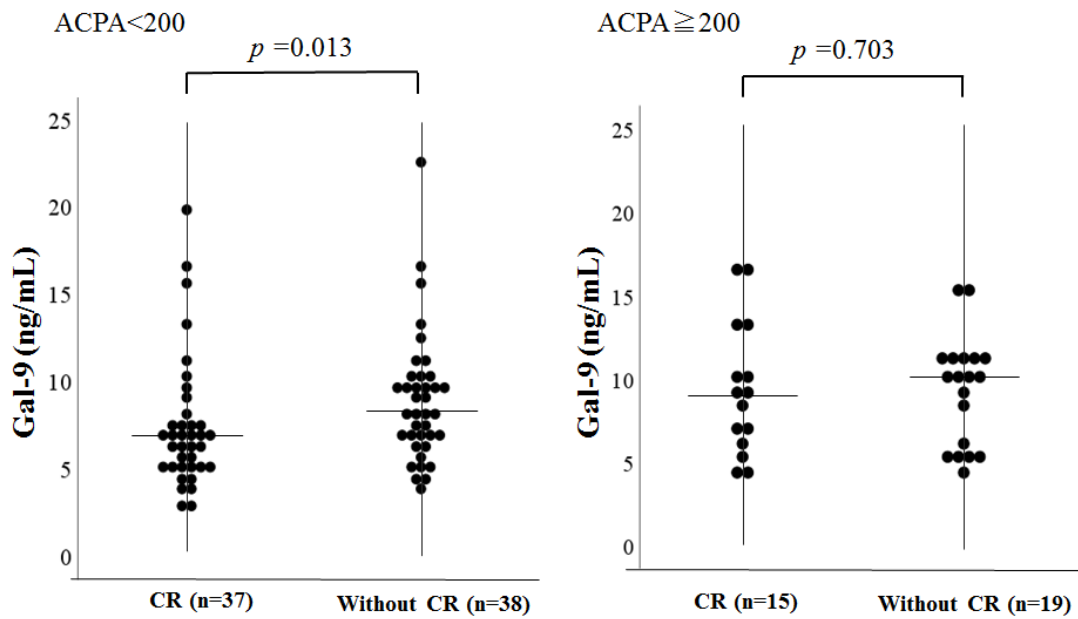


Figure 5. Serum levels of galectin-9 in RA patients with or without advanced joint damage. We compared serum levels of galectin-9 between RA patients with or without advanced joint damage (stage I versus stage II–IV).

A. Serum levels of galectin-9 were significantly higher in RA patients with advanced joint damage (stage II–IV) compared to those without advanced joint damage (stage I).

B. We compared serum levels of galectin-9 between RA patients with or without advanced joint damage (stage II–IV versus stage I) according to the ACPA titers. Serum levels of galectin-9 were significantly higher in patients with advanced joint damage (stage II–IV) compared to those without advanced joint damage (stage I) in RA patients with low titers of ACPA (< 200 U/ml), whereas there was no significant difference in serum levels of galectin-9 between patients with and without advanced joint damage in RA patients with high titers of ACPA (≥ 200 U/ml).

