福島県立医科大学 学術機関リポジトリ



Title	Current progress in genomics and targeted therapies for neurofibromatosis type 2
Author(s)	Hiruta, Ryo; Saito, Kiyoshi; Bakhit, Mudathir; Fujii, Masazumi
Citation	Fukushima Journal of Medical Science. 69(2): 95-103
Issue Date	2023
URL	http://ir.fmu.ac.jp/dspace/handle/123456789/2156
Rights	© 2023 The Fukushima Society of Medical Science. This article is licensed under a Creative Commons [Attribution-NonCommercial-ShareAlike 4.0 International] license.
DOI	10.5387/fms.2023-05
Text Version	publisher

This document is downloaded at: 2024-05-13T16:10:24Z

[Review]



Current progress in genomics and targeted therapies for neurofibromatosis type 2

Ryo Hiruta¹⁾, Kiyoshi Saito²⁾, Mudathir Bakhit¹⁾ and Masazumi Fujii¹⁾

¹⁾Department of Neurosurgery, Fukushima Medical University, Fukushima, Fukushima, Japan, ²⁾Department of Neurosurgery, Fukushima Rosai Hospital, Iwaki, Fukushima, Japan

(Received February 7, 2023, accepted June 6, 2023)

Abstract

Neurofibromatosis type 2 (NF2), a multiple neoplasia syndrome, is a manifestation of an impaired expression of the merlin protein, exerting inhibitory effects on cell proliferation signals due to abnormalities of the NF2 gene located on chromosome 22. About half of patients inherit a germline mutation from a parent, and nearly 60% of de novo NF2 patients are estimated to have somatic mosaicism. The development of technical methods to detect NF2 gene mutation, including targeted deep sequencing from multiple tissues, improved the diagnostic rate of mosaic NF2. With improved understanding of genetics and pathogenesis, the diagnostic criteria for NF2 were updated to assist in identifying and diagnosing NF2 at an earlier stage. The understanding of cell signaling pathways interacting with merlin has led to the development of molecular-targeted therapies. Currently, several translational studies are searching for possible therapeutic agents targeting VEGF or VEGF receptors. Bevacizumab, an anti-VEGF monoclonal antibody, is widely used in many clinical trials aiming for hearing improvement or tumor volume control. Currently, a randomized, doublemasked trial to assess bevacizumab is underway. In this randomized control trial, 12 other Japanese institutions joined the principal investigators in the clinical trial originating at Fukushima Medical University. In this review, we will be discussing the latest research developments regarding NF2 pathophysiology, including molecular biology, diagnosis, and novel therapeutics.

Key words: neurofibromatosis type 2, merlin, somatic mosaicism, bevacizumab

Introduction

Neurofibromatosis type 2 (NF2) is a tumorigenesis syndrome characterized by bilateral vestibular schwannoma (VS). The phenotypic expression of NF2 varies in severity, onset, and tumor expression, including intracranial and/or spinal schwannomas, intracranial meningiomas, and ependymomas. Furthermore, the mutation type or location in the *NF2* gene can influence tumor phenotypic variations.

In this review, we will outline clinical characteristics, elucidate the genetic mutations, and discuss recent advances in genetic mutation analysis methods. We will also describe the molecular features

of merlin, explore the relationship between genotype and phenotype, and provide an explanation of the diagnostic criteria. Finally, we will consider the potential for new therapeutic interventions and treatment modalities.

Epidemiology

The disorder is estimated to occur in one in 25,000-33,000 births^{1,2)}. Although NF2 patients vary in age of onset and clinical phenotype, this disease causes early death due to the critical anatomical location of tumors and the temporal and spatial multiplicity of tumors. NF2-related tumors develop in susceptible organs, including the nervous sys-

©2023 The Fukushima Society of Medical Science. This article is licensed under a Creative Commons [Attribution-NonCommercial-ShareAlike 4.0 International] license.

tem, eyes, and skin, deriving from cells that have lost wild-type (normal) function due to mutation of both alleles of the *NF2* gene. The average onset age of symptoms is 18-24 years, and the average age at death is 36 years²).

An analysis of the national NF2 registry of the Japanese Ministry of Health, Labour and Welfare, including 807 NF2 cases (44% male, 56% female) enrolled in the period between 2009 to 2013, showed that the ratio of NF2 patients with a family history of NF2 to those without was 1 to 2^{3} . The same study found that the onset age varied widely from less than five years to 80 years, with a median age of 24 years. Among patients, 42% had onset at 25 years or older and 45% had onset at less than 25 years (13% were unknown). The prognosis of NF2 differs between the two age-of-onset groups, with the younger group being worse. According to a nationwide survey using personal clinical records in Japan in 1999⁴⁾, the patients with onset at an age younger than 25 years showed overall 5-, 10-, and 20-year survival rates following diagnosis of 80%, 60%, and 28%, respectively. Corresponding survival rates for patients with onset older than 25 years were 100%, 87%, and $62\%^4$. The younger group is known as the Wishart type, while the older group is the Gardner type⁵⁾. The problem with either type of NF2 is the progressive deterioration of activity or quality of life due to multiple tumors in the central nervous system and peripheral nerves. Although the tumor itself is benign, the disease represents a long-term threat to life expectancy.

Clinical features

The nervous system, eyes, and skin of NF2 patients are susceptible to lesions. Although bilateral VS is characteristic, schwannomas often occur in other cranial, spinal, and peripheral nerves. Meningiomas and non-vestibular schwannomas are other intracranial lesions associated with NF2. These patients could have ocular lesions involving cataracts, retinal epitheliomas, and retinal hyperplasia. Cutaneous lesions also present frequently⁶⁾. It should be noted that "neurofibroma" might be present in NF2 patients but is quite rare. Historically, many have referred to this disease as neurofibromatosis, but this terminology is not representative of all NF2 clinical presentations. Thus, further consideration of the nomenclature for neurofibromatosis type 2 is necessary⁷.

A Japanese survey based on personal clinical records³⁾ found that almost 90% of the cases were

affected by vestibular schwannomas, the most frequent type of tumor in NF2, of which 81% were bilateral, and 6% were unilateral. Also, trigeminal schwannomas occurred bilaterally in 24% and unilaterally in 13% (unknown 17%). Intracranial meningiomas were found in 43% of the cases (unknown 11%). Hearing impairment was the most common symptom, with 61% of patients having severe hearing loss of >70 dB on at least one side of the ear. Other symptoms were spinal cord-related symptoms (48%), facial nerve palsy (34%), cerebellar ataxia, facial hypoesthesia, dysphagia or dysarthria, and others. In 58% of patients, progressive symptoms occurred, and only 3% experienced improvement.

Identifying NF2 gene

The first recorded case of bilateral VS was presented by Wishart in 1822⁸⁾. These diseases were previously thought to be Mendelian disorders that mainly affect the nervous system, with two clinically distinct types: type 1 neurofibromatosis, also known as von Recklinghausen neurofibromatosis, and type 2, also known as bilateral acoustic neurofibromatosis. Family studies involving nearly 100 members affected by neurofibromatosis type 2 suggested that this disorder was characterized by bilateral VS caused by a single mutant gene that was distinct from the mutation seen in neurofibromatosis type 1⁹⁾.

However, frequent loss of alleles on chromosome 22 was reported not only in NF2 but also in sporadic VS and meningiomas¹⁰⁾, suggesting that tumor proliferation was initiated by the inactivation of a tumor suppressor gene on the NF2 locus¹¹⁾. The specific loss of alleles on chromosome 22 was detected in various tumor types, including VS, other cranial neurofibromas, and meningiomas from patients with bilateral VS (i.e., NF2). Therefore, the common chromosome deletion mechanism was considered in different tumor types¹²⁾.

It was later discovered that NF2 patients carry a mutated NF2 gene on chromosome $22q12^{13,14)}$, which encodes merlin, an analog of the moesin-ezrin-radixin family of cytoskeleton-associated proteins¹⁵⁾.

Gene mutation analysis

Germline *NF2* pathogenic variants are identified in up to 90% of patients with a family history of NF2. The penetrance of NF2 is 100%, and those

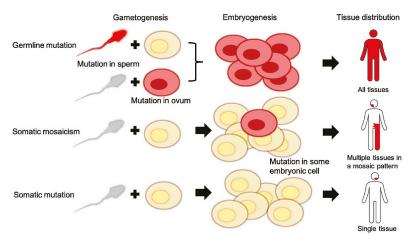


Fig. 1 Somatic mosaicism is a mosaic-like distribution of lesions resulting from genetic mutations in some embryonic cells during embryogenesis, which causes *de novo* NF2. Sporadic vestibular schwannoma follows somatic mutation.

with the *NF2* gene variant will develop the clinical disorder during their lifetime. On the other hand, more than 50% of NF2 patients have a *de novo NF2* gene variant without a family history, identifying *NF2* germline alterations are found in only 25%-60% of cases, and the remainder are thought to have somatic mosaicism¹⁶⁻¹⁸⁾. Because somatic mutations occur at the embryonic cell stage, a mosaic of different phenotypes expresses in normal tissues with wild-type *NF2* and tumor tissues with mutant $NF2^{19}$ (Figure 1). A study including >1,000 de novo NF2 patients estimated that nearly 60% of de novo cases have somatic mosaicism.

With regard to identifying the mutation of de novo NF2, sanger sequencing can identify point mutations or small-size mutations, while multiplex ligation-dependent probe amplification (MLPA) is suited to detect copy number alterations due to large deletions. However, it is challenging to identify NF2 variants with low variant allele frequency (VAF) in lymphocyte DNA. Thus, an analysis using these techniques may diagnose around 15% of cases as mosaic NF216). Even with next-generation sequencing (NGS), the genetic diagnostic rate was only between 20.2% to $23.5\%^{16,18,19)}$. If a case has a de novo somatic mosaicism, whether a variant could be detected in any cell lineage DNA depends on when the mutations developed in embryogenesis²⁰⁾. Thus, targeted deep sequencing of DNA from multiple tissues (blood, buccal mucosa, hair follicle, and tumor) can improve the diagnostic rate of mosaic NF2 by up to $37.7\%^{21}$.

Merlin protein

Merlin is related to the ezrin-radixin-moesin

(ERM) family of proteins that link plasma membranes and actin filaments. Merlin contains an N-terminal 4.1-ezrin-radixin-moesin (FERM) domain followed by a flexible coiled-coil domain and a carboxy-terminal hydrophilic tail (Figure 2). Merlin exists in two conformational forms, open and closed forms. Merlin switches from an active closed state to an inactive open state by phosphorylation at serine-518 on the carboxy-terminal domain²².

Merlin's function in cell signaling

Merlin exerts inhibitory effects on multiple receptor tyrosine kinases (RTK) such as the ErbB receptor, platelet-derived growth factor receptor (PDGFR), insulin-like growth factor 1 receptor (IG-F1R), and vascular endothelial growth factor receptor (VEGFR). This is confirmed by merlin-deficient schwannoma cell models strongly activating the downstream of these RTK, including oncogenic Ras/Raf/MEK/ERK and PI3K/AKT pathways^{23,24)}. Merlin also suppresses the activation of the mammalian target of rapamycin complex 1 (mTORC1) signaling in the mTOR pathway, which contributes to the control of cell survival and proliferation²⁵⁾, and the yes-associated protein (YAP) in the Hippo pathway regulating the cell proliferation and apoptosis 26 .

Cell signaling of activation and deactivation of merlin

One of the merlin activation mechanisms is contact inhibition, which regulates tissue growth when two cells contact each other. This important mechanism activates signals through membrane pro-

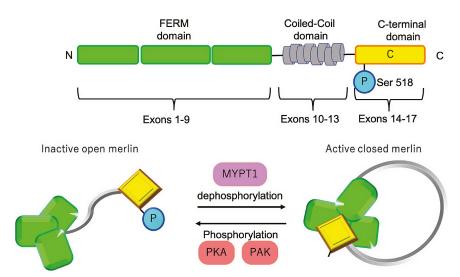


Fig. 2 Merlin consists of FERM, coiled-coil, and C-terminal domains. Dephosphorylation by MYPT1 closes the protein structure and activates merlin; phosphorylation by PKA or PAK opens the protein structure and deactivates merlin.

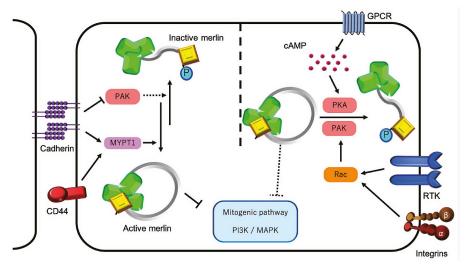


Fig. 3 (Left) Merlin is dephosphorylated (activated) by MYPT1, activated by cadherins and CD44, and suppresses cell proliferation signals. (Right) Tyrosine kinase receptors or integrins activate Rac and PAK, and G protein-coupled receptors signaling activate cyclic AMP and PKA, inducing phosphorylated (deactivated) merlin.

tein organization (i.e., CD44, epidermal growth factor receptor, and layilin), intercellular adhesion (i.e., cadherin), and cytoskeletal structures (i.e., spectrin and actin)²⁷⁾. The myosin phosphatase targeting subunit 1 (MYPT1) also dephosphorylates merlin protein into an active closed state²⁸⁾ (Figure 3).

Conversely, the mechanisms of merlin inactivation include the following. Proliferation signals initiated by membrane-anchored integrins and RTKs activate the signaling protein Rac, which activates p21-activated kinase (PAK), which in turn phosphorylates and inactivates merlin. In addition, a high concentration of cyclic AMP mediated by stimulation of G protein-coupled receptors (GPCRs) activates protein kinase A (PKA), which phosphory-

lates merlin²⁹⁾ (Figure 2).

Genotype-phenotype relationship

While phenotype and natural history of NF2 are often similar within the same family line, a different family line with other *NF2* mutations presents significant differences in phenotype, thought to be influenced by mutation type or location within the *NF2* gene³⁰. Patients with *de novo* NF2 also present phenotypic differences, suggesting a correlation with the extent of *NF2* VAFs²¹. The type of NF2 mutation is responsible for the number of lesions of the intracranial meningiomas, spinal cord, and peripheral nerves. If nonsense and frameshift muta-

tions occur in *NF2*, protein (merlin) expression is impaired, and the clinical symptoms are severe.

Meanwhile, missense mutations and in-frame deletions are known to result in mild symptoms because merlin could have a partial effect. An analysis of 268 patients with *NF2* abnormalities shows the truncated protein expression is associated with younger age at diagnosis and with a higher prevalence of meningiomas, spinal cord tumors, schwannomas rather than vestibular schwannomas, and skin tumors³¹⁾. The exon number where mutations occurred also affects the NF2 phenotype. An exon 14–15 mutation is associated with fewer clinical manifestations and meningiomas³²⁾, whereas truncating mutations in exons 2–13 present a worse prognosis than exon 14–15 mutations³⁰⁾.

The phenotypes are also different between germline mutation and somatic mosaicism. Mosaic NF2 patients show significantly lower VS growth rates, lower incidence rates of NF2-related lesions other than meningiomas, and higher age of onset. However, about half of mosaic NF2 patients have multiple large meningiomas, even with unilateral or mildly bilateral VS, suggesting that the phenotype is not always mild²¹⁾.

Clinical assessment for NF2 patients focused on hearing function. The more severe the NF2 mutation type, the more hearing is lost at a younger age³³). However, other reports showed that such a relation between the mutation type and hearing loss is not necessarily related to the tumor volume or growth rate ^{34,35}).

Differential diagnosis of NF2

The differential diagnosis of NF2 with significant clinical overlap includes schwannomatosis and familial syndrome of multiple meningiomas. Although schwannomatosis is similar to NF2 in presenting multiple schwannomas, it differs in the absence of bilateral VS. Schwannomatosis is associated with *SMARCB1* and *LZTR1* mutations located on chromosome 22. This disease has *NF2* somatic mutations in tumor tissue, suggesting that either *NF2* somatic mutations occur in each tumor or patients may have somatic mosaicism of *NF2*. It is reported that *SMARCB1* mutations do not cause VS, while *LZTR1* mutations do ³⁶⁾. Thus, revised diagnostic criteria for NF2 are proposed to include these genomic alterations¹⁷⁾.

Diagnostic criteria for NF2

The Manchester criteria developed in 1992 are the most widely used for NF2 diagnosis. With improved understanding of the genetics and pathogenesis of NF2, criteria have been revised to allow identification and diagnosis of NF2 at an earlier stage as a significant proportion of patients without bilateral VS^{17,36)} (Table 1).

Current status of NF2 drug discovery

Angiogenesis as a target of treatment

As discussed in NF2 gene function, merlin affects multiple pathways involved in cell growth, which can help to identify different therapeutic targets. Although VS is a benign tumor and grows relatively slowly, angiogenesis is required for the tumor to expand beyond a specific size^{37,38)}. Some angiogenesis-stimulating factors were identified, and among the most well-known is vascular endothelial growth factor (VEGF). A correlation has been found between the degree of VEGF expression and clinical parameters such as tumor growth, tumor volume, and microvessel density^{38,39)}. Currently, a number of translational research efforts for therapeutic targeting of VEGF or VEGF receptors have been conducted; the main ones are discussed below. Table 2 summarizes reports regarding other molecular-targeted therapies.

Bevacizumab

Bevacizumab, an anti-VEGF monoclonal anti-

Table 1. Diagnostic criteria for NF2

- 1. Bilateral vestibular schwannomas, or
- 2. FDR family history of NF2 and unilateral VS, or
- FDR family history of NF2 or unilateral VS and two
 of meningioma, cataract, glioma, neurofibroma,
 schwannoma, cerebral calcification (if UVS+, ≥2
 schwannomas only need negative LZTR1 test^a), or
- Multiple meningiomas (2 or more) and two of unilateral VS, cataract, glioma, neurofibroma, schwannoma, cerebral calcification, or
- Constitutional pathogenic NF2 gene variant in blood or identical in two tumors^b

NF2: neurofibromatosis type 2, FDR: first-degree relative, VS: vestibular schwannoma, UVS: unilateral vestibular schwannoma, LZTR1: leucine zipper-like transcription regulator 1

^aIncludes two of any tumor type, such as schwannoma ^bRequires molecular analysis

Table 2	Pocont	clinical	trials for	MF2 rol	2V bote
Table Z	Recent	Ciinicai	imais for	INFZ-rei	area vs

Drug	Author	Trial design and result
Lapatinib	Karajannis, et al ⁵⁰⁾	Phase II. 4/17 with >15% tumor size reduction 4/13 with >10 dB improved hearing
Imatinib	Stephanie, et al ⁵¹⁾	Case report. Prolonged VS progression for 4 months Due to adverse effects, changed to bevacizumab with prolonged progression for 8 months
Erlotinib	Plotkin, et al ⁵²⁾	Phase II. 11 patients with progressive VS No radiographic or hearing responses Median time to tumor progression was 9.2 months
Rapamycin	Giovannini, et al ⁵³⁾	A case presentation Arrested growth of progressive VS
Everolimus	Karajannis, et al ⁵⁴⁾	Phase II. 9 patients with progressive VS No radiographic or hearing responses.
Everolimus	Goutagny, et al ⁵⁵⁾	Phase II. No more than > 20% tumor size reduction 4/9 with progressive disease, 5/9 with stable disease Prolonged time to progression

body, is already approved for clinical use, including in Japan, for treating malignant gliomas and other carcinomas. Plotkin et al. investigated the effect of bevacizumab in NF2-related VS for the first time, showing a reduction in tumor volume in 9 of 10 patients and improved hearing in some cases⁴⁰⁾. Other reports showed the efficacy of bevacizumab in NF2-related VS⁴¹⁻⁴³⁾. Goutagny et al. reviewed the literature and reported that bevacizumab improved hearing and tumor shrinkage in more than 50% of progressive VS⁴⁴⁾. A study of bevacizumab administration in NF2-related VS in Japanese patients showed a maximum decrease in tumor volume to baseline occurring three months after receiving four doses of bevacizumab (5 mg/kg in two-week intervals)⁴⁵⁾. However, increasing the bevacizumab dose from 5 mg/kg every two weeks to 10 mg/kg was found to be ineffective⁴⁶⁾. The problem with bevacizumab therapy is the need for long-term administration for tumor control, which can result in side effects, including hypertension, proteinuria, and delayed wound healing.

Currently, a randomized, double-masked, place-bo-controlled, multicenter trial to assess bevacizum-ab's efficacy and safety in NF2 patients is ongoing in Japan, with sixty patients already included with the evaluation of hearing improvement in the affected ear as the primary objective⁴⁷⁾. This study is the first of its kind regarding the administration of bevacizumab in NF2 in a randomized controlled trial. It is also the most extensive prospective study of this rare and intractable disease, and the results are eagerly awaited. For this randomized control trial, 12 other Japanese institutions have joined the principal

investigators at Fukushima Medical University.

Peptide vaccine therapy for VEGF receptor

Another study investigating the development of peptide vaccines targeting the VEGF receptor (VEGFR) as a novel therapy is also underway in Japan. High VEGFR (VEGFR1 and VEGFR2) expression was detected on endothelial cells and tumor cells in NF2 schwannomas⁴⁸⁾. Thus, a subcutaneous injection of VEGFRs combined with a human leukocyte antigen (HLA) peptide vaccine could activate cytotoxic T lymphocytes (CTLs) specific for these and exert an antitumor effect. The CTLs attack tumor cells and blood vessels directly and are maintained as acquired immunity, expected to be a long-term effect. A Japanese clinical study showed that CTLs against both VEGFRs were induced in 6 out of 7 patients with eight doses of the peptide vaccine. Hearing improved in 40% of patients. Twenty-two out of 23 schwannoma lesions were reduced or had no change, but only one cystic schwannoma became enlarged. Furthermore, tumor specimens obtained from pre- and post-vaccination showed that the vaccination caused both loss of VEGFR expression and reduction of VEGF-A expression⁴⁹⁾.

Since this vaccine consists of HLA-A-binding peptides, some patients with different HLA-A types cannot induce an immune response to VEGFR. It is expected that the number of patients eligible for this kind of therapy could increase with the development of peptide vaccines mixed with certain common HLA types to compensate for the variations available in the population.

Conclusion and future prospects

Although NF2 is rare, it is still a life-threatening disorder with severe symptoms. There is some overlap with phenotypic-similar diseases (e.g., schwannomatosis or familial syndrome of multiple meningiomas); thus, genetic diagnosis will be essential. NF2 patients are offered either tumor debulking surgery or stereotactic radiotherapy according to their clinical symptoms. However, understanding the pathogenesis and employing genetic analysis can help diagnose NF2 early, especially the *de novo* types. Further clinical trials are still needed to assist in developing new management plans to improve the NF2 patient's quality of life.

Conflict of Interest

The authors are members of a research team conducting a phase 2 clinical trial to assess the therapeutic effect of bevacizumab in neurofibromatosis type 2. The trial is financed by the Japan Agency for Medical Research and Development (AMED) and the CHUGAI PHARMACEUTICAL COMPANY.

References

- Evans DG, Moran A, King A, Saeed S, Gurusinghe N, Ramsden R. Incidence of vestibular schwannoma and neurofibromatosis 2 in the North West of England over a 10-year period: higher incidence than previously thought. Otol Neurotol, 26: 93-97, 2005.
- 2. Evans DG. Neurofibromatosis type 2 (NF2): a clinical and molecular review. Orphanet J Rare Dis, 4: 16, 2009.
- 3. Iwatate K, Yokoo T, Iwatate E, Ichikawa M, Sato T, Fujii M, *et al.* Population Characteristics and Progressive Disability in Neurofibromatosis Type 2. World Neurosurg, **106**: 653-660, 2017.
- 4. Otsuka G, Saito K, Nagatani T, Yoshida J. Age at symptom onset and long-term survival in patients with neurofibromatosis Type 2. J Neurosurg, **99**: 480-483, 2003.
- Evans DG, Huson SM, Donnai D, Neary W, Blair V, Teare D, et al. A genetic study of type 2 neurofibromatosis in the United Kingdom. I. Prevalence, mutation rate, fitness, and confirmation of maternal transmission effect on severity. J Med Genet, 29: 841-846, 1992.
- 6. Asthagiri AR, Parry DM, Butman JA, Kim HJ, Tsilou ET, Zhuang Z, *et al.* Neurofibromatosis type 2. Lancet, **373**: 1974-1986, 2009.
- 7. Plotkin SR, Messiaen L, Legius E, Pancza P, Avery

- RA, Blakeley JO, *et al.* Updated diagnostic criteria and nomenclature for neurofibromatosis type 2 and schwannomatosis: An international consensus recommendation. Genet Med, 2022.
- 8. Wishart JH. Case of Tumours in the Skull, Dura Mater, and Brain. Edinb Med Surg J, **18**: 393-397, 1822.
- 9. Young DF, Eldridge R, Gardner WJ. Bilateral acoustic neuroma in a large kindred. JAMA, **214**: 347-353, 1970.
- 10. Zang KD. Cytological and cytogenetical studies on human meningioma. Cancer Genet Cytogenet, **6**: 249-274, 1982.
- 11. Seizinger BR, Martuza RL, Gusella JF. Loss of genes on chromosome 22 in tumorigenesis of human acoustic neuroma. Nature, **322**: 644-647, 1986.
- 12. Seizinger BR, Rouleau G, Ozelius LJ, Lane AH, St George-Hyslop P, Huson S, *et al.* Common pathogenetic mechanism for three tumor types in bilateral acoustic neurofibromatosis. Science, **236**: 317-319, 1987.
- 13. Rouleau GA, Merel P, Lutchman M, Sanson M, Zucman J, Marineau C, *et al.* Alteration in a new gene encoding a putative membrane-organizing protein causes neuro-fibromatosis type 2. Nature, **363**: 515-521, 1993.
- Wolff RK, Frazer KA, Jackler RK, Lanser MJ, Pitts LH, Cox DR. Analysis of chromosome 22 deletions in neurofibromatosis type 2-related tumors. Am J Hum Genet, 51: 478-485, 1992.
- Trofatter JA, MacCollin MM, Rutter JL, Murrell JR, Duyao MP, Parry DM, et al. A novel moesin-, ezrin-, radixin-like gene is a candidate for the neurofibromatosis 2 tumor suppressor. Cell, 72: 791-800, 1993.
- 16. Evans DG, Ramsden RT, Shenton A, Gokhale C, Bowers NL, Huson SM, et al. Mosaicism in neurofibromatosis type 2: an update of risk based on uni/bilaterality of vestibular schwannoma at presentation and sensitive mutation analysis including multiple ligation-dependent probe amplification. J Med Genet, 44: 424-428, 2007.
- 17. Evans DG, King AT, Bowers NL, Tobi S, Wallace AJ, Perry M, *et al.* Identifying the deficiencies of current diagnostic criteria for neurofibromatosis 2 using databases of 2777 individuals with molecular testing. Genet Med, **21**: 1525-1533, 2019.
- Louvrier C, Pasmant E, Briand-Suleau A, Cohen J, Nitschke P, Nectoux J, et al. Targeted next-generation sequencing for differential diagnosis of neurofibromatosis type 2, schwannomatosis, and meningiomatosis. Neuro Oncol, 20: 917-929, 2018.
- Evans DG, Hartley CL, Smith PT, King AT, Bowers NL, Tobi S, et al. Incidence of mosaicism in 1055 de novo NF2 cases: much higher than previ-

ous estimates with high utility of next-generation sequencing. Genet Med, **22**: 53-59, 2020.

- 20. D'Gama AM, Walsh CA. Somatic mosaicism and neurodevelopmental disease. Nat Neurosci, **21**: 1504-1514, 2018.
- 21. Teranishi Y, Miyawaki S, Hongo H, Dofuku S, Okano A, Takayanagi S, *et al.* Targeted deep sequencing of DNA from multiple tissue types improves the diagnostic rate and reveals a highly diverse phenotype of mosaic neurofibromatosis type 2. J Med Genet, **58**: 701-711, 2021.
- Shimizu T, Seto A, Maita N, Hamada K, Tsukita S, Tsukita S, et al. Structural basis for neurofibromatosis type 2. Crystal structure of the merlin FERM domain. J Biol Chem, 277: 10332-10336, 2002.
- Ammoun S, Flaiz C, Ristic N, Schuldt J, Hanemann CO. Dissecting and targeting the growth factor-dependent and growth factor-independent extracellular signal-regulated kinase pathway in human schwannoma. Cancer Res, 68: 5236-5245, 2008.
- 24. Neff BA, Voss SG, Schmitt WR, Driscoll CL, Link MJ, Beatty CW, *et al.* Inhibition of MEK pathway in vestibular schwannoma cell culture. Laryngoscope, **122**: 2269-2278, 2012.
- Lopez-Lago MA, Okada T, Murillo MM, Socci N, Giancotti FG. Loss of the tumor suppressor gene NF2, encoding merlin, constitutively activates integrin-dependent mTORC1 signaling. Mol Cell Biol, 29: 4235-4249, 2009.
- Zhang N, Bai H, David KK, Dong J, Zheng Y, Cai J, et al. The Merlin/NF2 tumor suppressor functions through the YAP oncoprotein to regulate tissue homeostasis in mammals. Dev Cell, 19: 27-38, 2010.
- de Vries M, van der Mey AG, Hogendoorn PC. Tumor Biology of Vestibular Schwannoma: A Review of Experimental Data on the Determinants of Tumor Genesis and Growth Characteristics. Otol Neurotol, 36: 1128-1136, 2015.
- 28. Jin H, Sperka T, Herrlich P, Morrison H. Tumorigenic transformation by CPI-17 through inhibition of a merlin phosphatase. Nature, **442**: 576-579, 2006.
- 29. Cooper J, Giancotti FG. Molecular insights into NF2/Merlin tumor suppressor function. FEBS Lett, **588**: 2743-2752, 2014.
- 30. Hexter A, Jones A, Joe H, Heap L, Smith MJ, Wallace AJ, *et al.* Clinical and molecular predictors of mortality in neurofibromatosis 2: a UK national analysis of 1192 patients. J Med Genet, **52**: 699-705, 2015.
- 31. Selvanathan SK, Shenton A, Ferner R, Wallace AJ, Huson SM, Ramsden RT, *et al.* Further genotype–phenotype correlations in neurofibromatosis 2. Clin Genet, **77**: 163-170, 2010.

- 32. Smith MJ, Higgs JE, Bowers NL, Halliday D, Paterson J, Gillespie J, *et al.* Cranial meningiomas in 411 neurofibromatosis type 2 (NF2) patients with proven gene mutations: clear positional effect of mutations, but absence of female severity effect on age at onset. J Med Genet, 48: 261-265, 2011.
- Halliday D, Emmanouil B, Pretorius P, MacKeith S, Painter S, Tomkins H, et al. Genetic Severity Score predicts clinical phenotype in NF2. J Med Genet, 54: 657-664, 2017.
- 34. Masuda A, Fisher LM, Oppenheimer ML, Iqbal Z, Slattery WH, Natural History C. Hearing changes after diagnosis in neurofibromatosis type 2. Otol Neurotol, **25**: 150-154, 2004.
- 35. Graamans K, Van Dijk JE, Janssen LW. Hearing deterioration in patients with a non-growing vestibular schwannoma. Acta Otolaryngol, **123**: 51-54. 2003.
- Smith MJ, Bowers NL, Bulman M, Gokhale C, Wallace AJ, King AT, et al. Revisiting neurofibromatosis type 2 diagnostic criteria to exclude LZTR1-related schwannomatosis. Neurology, 88: 87-92, 2017.
- 37. Hanahan D, Folkman J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. Cell, 86: 353-364, 1996.
- 38. Caye-Thomasen P, Werther K, Nalla A, Bog-Hansen TC, Nielsen HJ, Stangerup SE, *et al.* VEGF and VEGF receptor-1 concentration in vestibular schwannoma homogenates correlates to tumor growth rate. Otol Neurotol, **26**: 98-101, 2005.
- 39. Koutsimpelas D, Stripf T, Heinrich UR, Mann WJ, Brieger J. Expression of vascular endothelial growth factor and basic fibroblast growth factor in sporadic vestibular schwannomas correlates to growth characteristics. Otol Neurotol, **28**: 1094-1099, 2007.
- 40. Plotkin SR, Stemmer-Rachamimov AO, Barker FG, 2nd, Halpin C, Padera TP, Tyrrell A, *et al.* Hearing improvement after bevacizumab in patients with neurofibromatosis type 2. N Engl J Med, **361**: 358-367, 2009.
- 41. Plotkin SR, Merker VL, Halpin C, Jennings D, McKenna MJ, Harris GJ, *et al.* Bevacizumab for progressive vestibular schwannoma in neurofibromatosis type 2: a retrospective review of 31 patients. Otol Neurotol, **33**: 1046-1052, 2012.
- 42. Eminowicz GK, Raman R, Conibear J, Plowman PN. Bevacizumab treatment for vestibular schwannomas in neurofibromatosis type two: report of two cases, including responses after prior gamma knife and vascular endothelial growth factor inhibition therapy. J Laryngol Otol, 126: 79-82, 2012.
- 43. Mautner VF, Nguyen R, Kutta H, Fuensterer C, Bokemeyer C, Hagel C, et al. Bevacizumab in-

- duces regression of vestibular schwannomas in patients with neurofibromatosis type 2. Neuro Oncol, 12: 14-18, 2010.
- Goutagny S, Kalamarides M. Medical treatment in neurofibromatosis type 2. Review of the literature and presentation of clinical reports. Neurochirurgie, 64: 370-374, 2018.
- 45. Fujii M, Ichikawa M, Iwatate K, Bakhit M, Yamada M, Kuromi Y, *et al.* Bevacizumab Therapy of Neurofibromatosis Type 2 Associated Vestibular Schwannoma in Japanese Patients. Neurol Med Chir (Tokyo), **60**: 75-82, 2020.
- 46. Plotkin SR, Duda DG, Muzikansky A, Allen J, Blakeley J, Rosser T, et al. Multicenter, Prospective, Phase II and Biomarker Study of High-Dose Bevacizumab as Induction Therapy in Patients With Neurofibromatosis Type 2 and Progressive Vestibular Schwannoma. J Clin Oncol, 37: 3446-3454, 2019.
- 47. Fujii M, Kobayakawa M, Saito K, Inano A, Morita A, Hasegawa M, et al. Rationale and Design of Beat-NF2 Trial: A Clinical Trial to Assess the Efficacy and Safety of Bevacizumab in Patients with Neurofibromatosis Type 2 Related Vestibular Schwannoma. Curr Oncol, 28: 726-739, 2021.
- Tamura R, Morimoto Y, Sato M, Kuranari Y, Oishi Y, Kosugi K, et al. Difference in the hypoxic immunosuppressive microenvironment of patients with neurofibromatosis type 2 schwannomas and sporadic schwannomas. J Neurooncol, 146: 265-273 2020
- 49. Tamura R, Fujioka M, Morimoto Y, Ohara K, Kosu-

- gi K, Oishi Y, *et al.* A VEGF receptor vaccine demonstrates preliminary efficacy in neurofibromatosis type 2. Nat Commun, **10**: 5758, 2019.
- 50. Karajannis MA, Legault G, Hagiwara M, Ballas MS, Brown K, Nusbaum AO, et al. Phase II trial of lapatinib in adult and pediatric patients with neurofibromatosis type 2 and progressive vestibular schwannomas. Neuro Oncol, 14: 1163-1170, 2012.
- 51. Lim S, de Souza P. Imatinib in neurofibromatosis type 2. BMJ Case Rep, **2013**: 2013.
- 52. Plotkin SR, Halpin C, McKenna MJ, Loeffler JS, Batchelor TT, Barker FG, 2nd. Erlotinib for progressive vestibular schwannoma in neurofibromatosis 2 patients. Otol Neurotol, **31**: 1135-1143, 2010.
- 53. Giovannini M, Bonne NX, Vitte J, Chareyre F, Tanaka K, Adams R, et al. mTORC1 inhibition delays growth of neurofibromatosis type 2 schwannoma. Neuro Oncol, 16: 493-504, 2014.
- 54. Karajannis MA, Legault G, Hagiwara M, Giancotti FG, Filatov A, Derman A, *et al.* Phase II study of everolimus in children and adults with neurofibromatosis type 2 and progressive vestibular schwannomas. Neuro Oncol, **16**: 292-297, 2014.
- 55. Goutagny S, Raymond E, Esposito-Farese M, Trunet S, Mawrin C, Bernardeschi D, et al. Phase II study of mTORC1 inhibition by everolimus in neurofibromatosis type 2 patients with growing vestibular schwannomas. J Neurooncol, 122: 313-320, 2015